



Available Online at

<http://www.ijcpa.in>

IJCPA, 2014; 1(3): 130-140

International Journal of  
CHEMICAL AND PHARMACEUTICAL  
ANALYSIS

ISSN: 2348-0726

Review Article

Immunomodulatory and Therapeutic Potential of CPG and Probiotic DNA – A Review

Aruna Bhatia<sup>1\*</sup>, Mansimran Kaur Randhawa<sup>2</sup>, Manpreet Kaur<sup>3</sup>

<sup>1</sup>Immunology and Immunotechnology Lab, Department of Biotechnology, Punjabi University, Patiala – 147002, Punjab, India

<sup>2</sup>Department of Biopharma Sciences, Chandigarh University, Gharuan, Mohali, Punjab, India

<sup>3</sup>Immunology and Immunotechnology Lab, Department of Biotechnology, Punjabi University, Patiala, India

Received: 12 June 2014 / Revised: 29 June 2014 / Accepted: 30 June 2014/ Online publication: 1 July 2014

**ABSTRACT**

The in depth studies on the immune response shows its application in diseases other than infections. There are hardly any chemical based drugs which are given as immunomodulators. Most of the disease treatment drugs available in the market may have side effects and are costly. Hence, alternatively the immunotherapy can be employed to manage immune related diseases by the use of natural agents like probiotics. Moreover, the literature shows that prokaryotic DNA can activate immune response and has potential therapeutic applications. This immune efficacy of DNA is due to the presence of unmethylated DNA i.e. CpG motifs (regions of DNA where Cytosine and Guanine are separated by only one phosphate) in prokaryotes. Hence, the various applications of prokaryotic as well as probiotic DNA are being explored in the treatment of immune related disorders. The status of Probiotic DNA and synthetic oligodeoxynucleotides containing CpG motifs (CpG-ODNs) from various microorganisms are reviewed here, with reference to some key examples where these have succeeded in prevention and treatment of diseases.

**Keywords:** Probiotics, DNA, CpG-ODN, Immunomodulatory Therapy, Cytokines

**1. INTRODUCTION**

The immune system is a versatile defence system and has evolved to protect animals, humans and other living beings from invading pathogens and infectious diseases. The increase in knowledge about the immune response mechanisms unveils the relationship of immune response and related disorders. This made clear that besides immunological disorders, many other diseases can be modified by modulating Immune response. The rise in infectious diseases as well as non infectious immune related disorders demand the need for the development of

efficient immunomodulators. Various medications including chemical drugs as well as natural therapies have been tested for curing such diseases. The chemical based drugs employed to cure these diseases may have the side effects and are costly. Moreover, consumer awareness about the harmful effects of chemical drugs raised a need to search for natural/alternative therapies for the treatment of diseases. Immunotherapy is one of the alternative ways of modification of diseases. Probiotics, the beneficial bacteria have been proved to have immunostimulating efficacy. These bacteria have GRAS (Generally Regarded As Safe) status, which made them most consumer acceptable alternative therapeutic agent. The advances in techniques in molecular biology, immunology and microbiology lead towards finding that not only the probiotic

**\*Corresponding Author:**

Dr. Aruna Bhatia  
Professor, Immunology and Immunotechnology Lab.,  
Department of Biotechnology, Punjabi University, Patiala, India  
Contact No. +91- 9878263077  
Email: [aruna\\_bhatia@rediffmail.com](mailto:aruna_bhatia@rediffmail.com)

but their DNA alone also is capable of immune activation. Preliminary experiments revealed that probiotic DNA is a better immune enhancer than its whole cell. This immunomodulating potential is due to the presence of CpG motifs in the DNA of probiotic microorganism, being a prokaryote.

## 2. PROBIOTICS

Probiotics are defined as viable microorganisms, sufficient amounts of which reach the intestine in an active state and thus exert positive health effects. Probiotics are live bacteria which are intended to colonize the large intestine and confer physiological health benefits to the host, when administered in adequate amounts and they promote or support a beneficial balance of microbes to live in the gastrointestinal tract<sup>1</sup>. The growing awareness of relationship between diet and health and the side effects of allopathic medication has led to an increase in the demand for the food products that support health beyond providing nutrition and act as an alternative source of medicine. Hence, the use of probiotics appear as a new tool for the treatment of diseases<sup>2</sup>. In addition, the bioefficacy of the probiotics can be enhanced by the use of prebiotics. Prebiotic is a non-digestible food ingredient that beneficially affects the host by selectively stimulating the growth and/or activity of one or a limited number of bacteria in the colon, and thus improves host health. Thus, the synergistic effect of prebiotics and probiotics as synbiotics is useful in improving the health of the gut. In addition, prebiotics also have following health effects<sup>3</sup>:

- Improvement of blood glucose responses.
- Relieve acute and chronic constipation.
- Inhibition of secondary bile acid formation.
- Treatment of salmonellosis.
- Enhancement of mineral absorption.

### 2.1 Specific roles and benefits of probiotics in the body

- Prevention of genital and urinary tract infections<sup>4,5</sup>
- Nutrient synthesis and bioavailability<sup>6</sup>
- Prevention of infection by Pathogenic bacteria<sup>7</sup>
- Alleviation of the symptoms of Rheumatoid Arthritis<sup>8,9,10</sup>
- Establishment of a healthy microbiota in preterm infants<sup>11</sup>
- Increased cellular immunity (e.g. increased natural killer cell activity)<sup>12</sup>

- Alleviation of Lactose maldigestion<sup>13</sup>
- Anti- allergic potential<sup>14,15</sup>
- Immune tolerance<sup>16</sup>
- Lowered levels of cold and influenza-like symptom in children<sup>17</sup>
- Lowered incidence of diarrhoea<sup>18</sup>
- Immunostimulatory capability<sup>19</sup>
- Anti-carcinogenic activity<sup>20</sup>
- Lowering of blood cholesterol levels<sup>21,22</sup>
- Hypoglycemic activity<sup>23</sup>
- Reduction of pro-inflammatory cytokine expression<sup>24</sup>

Due to the above mentioned properties, probiotic bacteria lead the researchers to place them as one of the food supplements and fortifying agents. Food sources of probiotic bacteria also include fermented dairy products like yoghurt, milk with live culture and also sold as kefir, koumiss and yakult, etc.

### 2.2 Mechanism of action of Probiotics

Besides showing their antimicrobial activity directly by releasing bacteriocins, the mechanism by which probiotics exert their bioactivity in gut include their effect on immune system of host as has been given in fig.1 by Shida and Nanno<sup>25</sup>

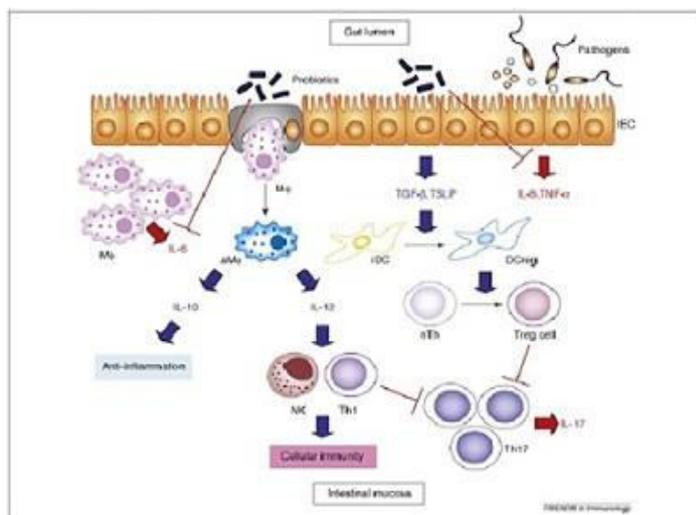


Fig.1: Mechanism of Probiotics in Gut

On exposure to pathogen microorganisms, intestinal epithelial cells (IEC) produce pro-inflammatory mediators such as IL8 and TNFα. The production of these cytokines is inhibited by probiotics and anti-inflammatory mediators such as TGF-β are produced, which can stimulate the differentiation of immature dendritic cells (DCs), followed by the induction of regulatory T cells (Treg). These cells exercise an anti-inflammatory function

by controlling Th1, Th2 and probably Th17 cells. The macrophages (M) produce large amounts of IL6 in the inflamed mucosa. The probiotics may determine the reduction of this production and the increase of the production of IL10. The intestine usually produces small quantities of IL12, but probiotics have the potential to increase this production.

### 2.3 Cellular components of Probiotic Bacteria and their effect on host immunity

It has been reported that cellular components of Lactic acid Bacteria (LAB) as well as live bacteria<sup>26</sup> and inactivated or heat killed bacteria<sup>27,28</sup> could exert immunoregulating effects. Other components of bacteria such as peptidoglycan & teichoic acid<sup>29</sup>; cell surface protein<sup>30,31</sup>; exopolysaccharides<sup>32</sup>, etc provide immunoregulatory effects.

Bacterial DNA is also being explored as an immune enhancer<sup>33</sup>. The vertebrate immune system recognizes 'CpG motifs' of probiotic DNA as foreign and trigger protective immune responses which are strongly Th1-based. A study conducted by Krieg et al.<sup>34</sup> and Weeratna et al.<sup>35</sup>, proved that bacterial DNA acts as a strong vaccine adjuvant, for inducing humoral immunity. These cellular components are known as pathogen associated molecular patterns (PAMPs) and they exert their effect by interacting with Pattern Recognition Receptors (PRRs) of immunocompetent cells<sup>36,37</sup> because, PAMPs are considered as the ligands of PRRs.

Probiotic DNA has the following benefits over whole cell bacteria-

- Amelioration of bioactivity by the use of DNA is more than whole cell probiotic bacteria<sup>38</sup>.
- Whole cell probiotics are not useful in certain immunocompromised persons and the persons who are already extremely ill<sup>39</sup>.

### 2.4 Health effects of Probiotic DNA

Over the past few years, there has been a dramatic increase in understanding the molecular and cellular effects of CpG DNA and its effects *in vivo* in animal models. Studies till date suggest that CpG DNA can be used in the treatment of a variety of diseases including infection, allergy and cancer.

The immunodulatory effects of probiotic DNA has been observed by Lammers et al.<sup>40</sup>. In this study, peripheral blood mononuclear cells (PBMC) from healthy donors were incubated with pure DNA of eight probiotic strains and with total bacterial DNA from human faeces collected before and after probiotic ingestion. It was concluded that *Bifidobacterium* genomic DNA induced secretion of the anti-inflammatory interleukin-10 by PBMC. Iliev et al.<sup>33</sup> found that DNA from *Lactobacillus rhamnosus* GG (LGG) has a strong stimulatory effect on murine B cell proliferation. It was investigated that the whole genomic DNA isolated from LGG applied onto the murine splenic B cell cultures had mitogenic activity and that the chromosomal DNA from the strain at a concentration 50 µg/ml raised strong proliferation response in murine B cells.

Rachmilewitz et al.<sup>41</sup> demonstrated that the immunostimulatory activity of the genomic DNA together with other factors of probiotic activity is involved in the mechanism of protection of gut by probiotic bacteria. Probiotic bacteria DNA can also suppress systemic inflammatory responses to pathogenic bacterial DNA<sup>42,43</sup>. Moreover, many CpG motifs have been identified in probiotic bacteria<sup>44</sup>. Takahashi et al.<sup>45</sup> proved the anti-allergic activity of probiotic DNA. It was investigated that ODN (oligodeoxynucleotides) BL07 of *Bifidobacterium longum* BB536 strain, significantly inhibited secretion of IgE in B cells in the presence of IL-4 and anti-CD40.

The experiments were conducted by Medina et al.<sup>46</sup> to evaluate the ability of different strains of *Bifidobacterium longum* along with their DNA to induce cytokine production by peripheral blood mononuclear cells (PBMCs). It was proved that live cells of all *B. longum* strains stimulated regulatory cytokine interleukin (IL)-10 and proinflammatory cytokine tumour necrosis factor (TNF-α) production. Genomic DNA of some strains stimulated the production of the Th1 and pro-inflammatory cytokines, interferon (IFN)-γ and TNF-α, but not that of IL-10. Ghadimi et al.<sup>14</sup> experimentally proved that the Gram-positive (probiotic) *Lactobacilli* and *Bifidobacteria* tested, as well as their genomic DNA, dose-dependently modulated the TH1/TH2 response to allergens. DNA seemed to contribute to about 50% of the immunomodulatory effects exerted by live bacteria.

Zhong et al.<sup>47</sup> investigated the effect of living probiotics, probiotic DNA and the CpG-ODNs on both immune response and intestinal barrier function in ovalbumin-sensitized rat and the underlying mechanisms. It was concluded that the probiotic genomic DNA and CpG-ODN was comparable with living probiotics in preventing food allergic response by immune modulation and intestinal barrier function enhancement, and the activation of TLR9/NF- $\kappa$ B signal pathway might be involved in this process.

In our own lab, Randhawa et al.<sup>38</sup> conducted a study to compare the *in vivo* efficacy of 3 probiotic strains having maximum (*Lactobacillus delbrueckii* 405), moderate (*Lactobacillus brevis* 403), minimum (*Bifidobacterium bifidium* BD 234) immune activity and their isolated DNA (DNA LB 405, DNA 403, DNA Bif 234) on the basis of evoking the immune response in Swiss albino mice. The results demonstrated that a substantial augmentation in immune efficacy was observed in the animals receiving genomic DNA over the group receiving viable bacteria. Hence, it was concluded in their thesis that to get the immune effects, it is not necessary to give whole bacterial cell in the host. Instead, bacterial DNA of immunoactive probiotic can be used as a safe immunobiotherapeutic agent (anti-diabetic, anti-cholesteremic, immunorestorer) even in immunocompromised host. The immunostimulating activity of DNA depends on the presence of CpG motifs.

Various researchers explored the significance and application of CpG-ODNs in modulating immune system as well as preventing and curing various immune related diseases.

### 3. CpG DNA

Current knowledge indicates that prokaryotic DNA can activate immune response and has potential therapeutic application. The initial discovery of this phenomenon was reported by Tokunaga et al.<sup>48</sup>, who found out that *in vivo* DNA purified from *Mycobacterium* sp. (but not DNA from vertebrates) activated Natural Killer (NK) cells fostered the release of IFN- $\gamma$  by these cells and caused tumor regression.

Krieg et al.<sup>34</sup> confirmed that bacterial DNA was immunostimulatory in a way that mammalian DNA was not, and identified a sequence pattern of bases, or motif, responsible for

this property. Indeed, methylation of cytosine residues in the bacterial DNA or in the corresponding oligodeoxynucleotides destroyed their immunostimulatory activities<sup>34</sup>. CpG motifs are present in significantly greater frequency in bacterial DNA than vertebrate DNA. Specifically, bacterial DNA is thought to activate inflammatory cells because of its high content unmethylated CpG dinucleotides<sup>49</sup>.

#### 3.1 Mechanism of action of CpG DNA

Bacterial DNA and CpG-ODNs activate Antigen Presenting Cells (APCs) such as macrophages and dendritic cells. CpG-ODNs interact with CpG binding protein at the cell surface. After endocytosis and recognition by Toll like receptor 9 (TLR-9), (one of the members of the Toll family of innate immune receptors, that detect molecular patterns related to intracellular pathogens), TLR9 + CpG then interact with myeloid differentiation primary response gene 88 (MYD88) and the inhibitor of nuclear factor  $\kappa$ B- (NF- $\kappa$ B) kinase (IKK) complexes and the complex is translocated to the cell nucleus. The location of TLR-9 within the endosome provides both rapid and specific detection of pathogenic DNA and protection against activation by non-pathogenic "self" DNA<sup>50</sup>. Once in the cell nucleus, the CpG-ODNs activate NF- $\kappa$ B and initiate the induction of cytokines and chemokines (Fig. 2) by Fonseca and Kline<sup>51</sup>.

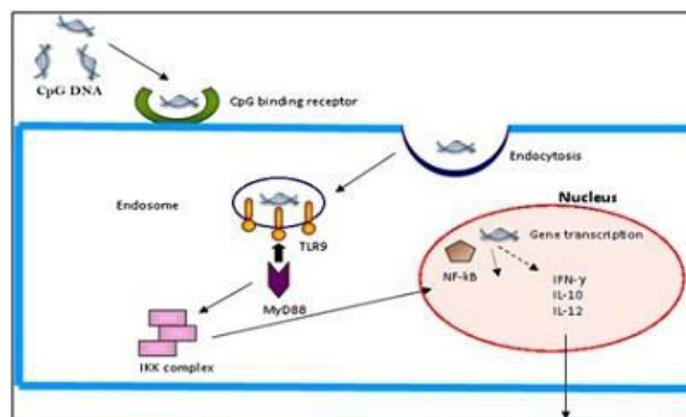


Fig. 2: Mode of action of CpG DNA

CpG-DNA also induces the monocytes and macrophages to produce inflammatory cytokines such as IL-6, IL-12, IFN- $\alpha$ , IFN- $\beta$ , TNF- $\alpha$ , IL-1 $\beta$ , IL-18, which mediate ADCC (Antibody Dependent Cell Cytotoxicity), express inducible nitric oxide synthase and promotes lytic activity of NK cells and the secretion of IFN- $\gamma$ <sup>52,53</sup>. As shown in fig. 3, by Agrawal and Kandimalla<sup>54</sup>, CpG DNA

exhibit several immunological effects that has led to their use as therapeutic agents and adjuvants for various diseases.

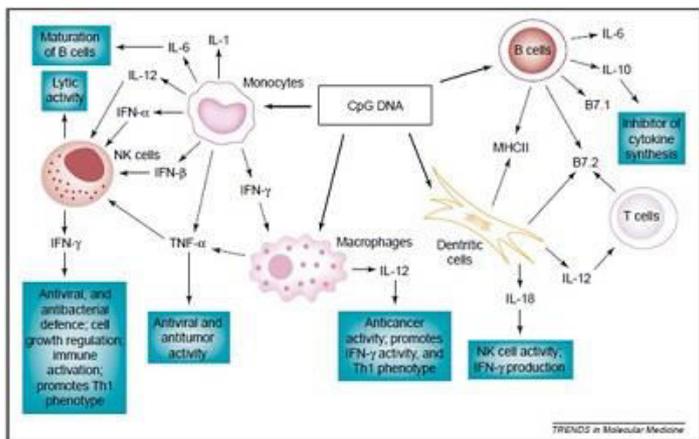


Fig. 3: Effects of CpG DNA on host immune cells

### 3.2 Immunotherapeutic applications of CpG DNA

The immune efficacy of CpG-ODNs resulted in the testing of their therapeutic potential. The potential applications CpG-ODNs (as mentioned in fig. 4) are as follows:

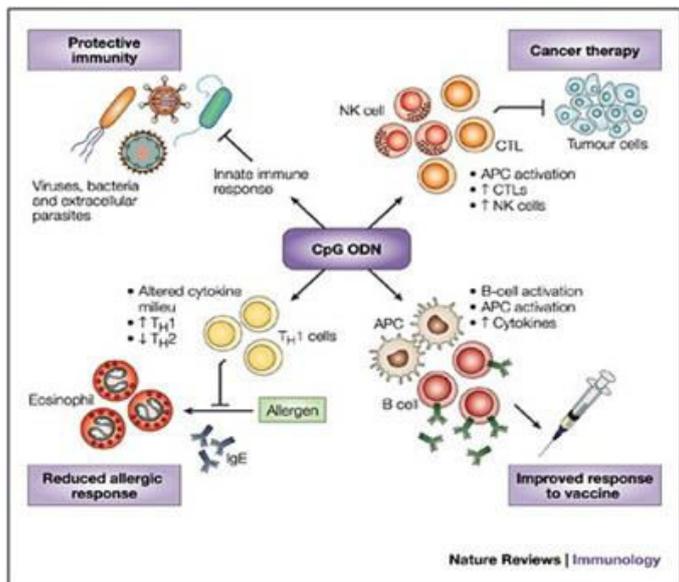


Fig. 4: Potential therapeutic uses of CpG oligodeoxynucleotides<sup>55</sup>

#### 3.2.1 Protection against infectious diseases

Krieg et al.<sup>56</sup> experimentally proved that when mice were injected with bacterial DNA or CpG-ODNs, a rapid production of IL-12 and IFN- $\gamma$  was noticed. The serum levels of IL-12 remain increased for at least 8 days after a single injection of CpG-ODNs, but IFN- $\gamma$  levels returned to baseline within 24 h. It was concluded that this Th1-like cytokine response to CpG motifs induces a state of resistance to infection by *Listeria*

*monocytogenes* in susceptible specific pathogen-free BALB/c mice. Gomis et al.<sup>57</sup> observed anti-infectious effect of DNA containing CpG motifs, when chickens were shown to be protected by infection from *Escherichia coli* due to the provision of intramuscular or subcutaneous injection doses to chickens prior to *E.coli* challenge. The study conducted by Jørgensen et al.<sup>58</sup> suggested that CpG DNA induces antiviral protection in Atlantic salmon fish.

Huang et al.<sup>59</sup> demonstrated that bacterial DNA plays an important role in the macrophage response to a heat killed pathogen *Brucella abortus*. Talati et al.<sup>60</sup> also reported the role of bacterial DNA/TLR9 pathway in implicating early host defense against *Streptococcus pneumoniae*. Zhu et al.<sup>61</sup> proved protective effect of CpG DNA against mastitis induced by *Escherichia coli* infection in rat model. Easton et al.<sup>62</sup> conducted a study and proved that intranasal delivery of CpG oligodeoxynucleotides provided significant protection to susceptible BALB/c mice against the Gram-negative bacterium *Burkholderia pseudomallei*, which is the causative agent of melioidosis. Judy et al.<sup>63</sup> investigated that the prophylactic application of CpG oligonucleotides augments the early host response and confers protection in acute melioidosis.

#### 3.2.2 Development of Vaccine/Vaccine adjuvant

McCluskie and Krieg<sup>64</sup> reviewed that CpG-ODNs greatly enhance the speed and strength of the immune responses if used as vaccine adjuvants. Krieg et al.<sup>34</sup> proposed the mechanisms contributing to the strong adjuvant activity of the CpG motifs for inducing humoral immunity may include:

- i) Synergy between TLR9 and the B cell receptor (BCR) preferentially stimulating antigen specific B-cells.
- ii) Inhibition of B-cell apoptosis improving B-cell survival<sup>65</sup>.
- iii) Enhanced IgG class switch DNA recombination which may enhance the maturation of the immune response<sup>66,67</sup>.

Kojima et al.<sup>68</sup> observed adjuvant effects of multi CpG motifs on an HIV DNA vaccine. The results suggested that the immunogenicity of DNA vaccines can be augmented by the addition of CpG motifs. Cooper et al.<sup>69</sup> evaluated CpG DNA in human clinical trials as adjuvants for Hepatitis B surface antigen either in combination with alum or alone. Sugai et al.<sup>70</sup> proved that a CpG-ODN acts as an efficient adjuvant counterbalancing

the Th1/Th2 immune response in Diphtheria-Tetanus-Pertussis (DPT) vaccine.

Maurer et al.<sup>71</sup> examined that CpG-ODN boost cytokine and stimulatory molecule expression on murine bone marrow-derived dendritic (mBMDC). It was concluded that CpG-DNA acts as potent adjuvant for vaccination therapies. Zhao et al.<sup>72</sup> investigated the protective response against *Treponema pallidum* (Tp) infection of a DNA vaccine enhanced by an adjuvant CpG-ODN. It was proved that CpG-ODN stimulated the secretion of IL-2 and IFN- $\gamma$  and protective effect was observed due to decrease in the incidence of Tp-infection of skin lesions ulceration lesions. Wang et al.<sup>73</sup> evaluated CpG-ODN for its immunostimulatory activity in mice which were vaccinated with recombinant outer membrane protein P56 (rP56 $\Delta$ ) of *Orientia tsutsugamushi*. It was concluded by this study that CpG-ODN adjuvant could help in enhancing the rP56 $\Delta$  immune responses, elicit memory immune response (IgM antibody) quickly and shorten its vaccination schedule.

### 3.2.3 Antiallergic potential

Horner et al.<sup>74</sup> conducted *in vitro* study with PBMCs from both nonatopic and atopic donors to characterize the antiallergic potential of phosphorothioate ISS-ODN in human subjects. This study demonstrated that phosphorothioate ISS-ODN elicits an innate immune response by PBMCs, which inhibits IL-4-dependent IgE synthesis. In addition, ISS-ODN also stimulated the production of IgM, IgG, and IgA.

Kitagaki et al.<sup>75</sup> quoted that the oral administration of CpG-DNA can protect against eosinophilic airway inflammation in murine model of asthma in association with reduction of antigen specific IgE in a dose dependent manner. CpG-ODNs may be useful as a component of oral immunotherapy to promote tolerance in established asthma. Suzuki et al.<sup>76</sup> demonstrated, for the first time, that immunotherapy with CpG DNA conjugated with a T-cell peptide is useful in preventing and treating allergic conditions. Xu et al.<sup>77</sup> proved the ability of immunomodulatory CpG-ODN, which are potent inducers of Th1 cytokines to prevent allergic symptoms in mice immunized and sensitized with allergen. Ashino et al.<sup>78</sup> investigated that co-administration of CpG-ODNs with an antigen prevented airway

eosinophilia and IgE production as well as bronchial hyperreactivity in a murine model of asthma.

Miyazaki et al.<sup>79</sup> investigated the suppressive mechanism of CpG treatment in allergic conjunctivitis induced experimental mice. It was concluded that administration of the CpG-ODN induced significant splenomegaly and adoptive transfer of the splenocytes isolated from CpG-treated mice conferred resistance to inflammation in allergen-induced recipient mice. Farrokhi et al.<sup>80</sup> performed a study to evaluate the potential effects of Co-administration of *Chenopodium album* allergens and CpG-ODN (CpG/Ch.a) in the immune deviation from Th2 to Th1 responses in case of Allergic Rhinitis (AR) patients. Several cytokines such as IFN- $\gamma$ , IL-4, IL-13, IL-10 and Ch.a specific IgE were measured in culture supernatants of PBMCs of patients. This study suggested that co-administration of CpG/Ch.a is effective *in vitro* in suppression of Th2 and stimulation of Th1 cytokine production. Kaburaki et al.<sup>81</sup> suggested that Japanese cedar pollen allergen (Cry j 1) conjugated with CpG-ODNs immunostimulation can induce Cry j1 specific Th1 responses, thereby inhibiting IgE response to the pollen allergen. Hence, CpG-ODN may be an effective novel method of inducing protection against atopic disorders.

### 3.2.4 Antitumor immunization

Weeratna et al.<sup>35</sup> reported that CpG-ODNs can be used successfully in tumor immunotherapy in animal models as well as in human clinical trials. Preliminary studies in several animal models of cancer suggest that CpG DNA have the ability to induce tumor regression by activating innate immunity, enhancing antibody dependent cellular cytotoxicity and elicit a specific, protective immune response<sup>82</sup>. Cornet et al.<sup>83</sup> proved that CpG DNA activate dendritic cells *in vivo* and induce a functional and protective immunity. Study of the CD8 response obtained after antigenic challenge suggested that a functional memory response is induced upon vaccination with CpG-ODN. Thus, MHC class I – restricted epitope in combination with CpG DNA is a promising and rather simple cancer formulation. Vaccination with CpG-conjugated tumor cells induces the expansion of tumor-specific cytotoxic T lymphocytes (CTL) that reduce the growth of established tumors and prevents their metastatic spread<sup>84</sup>. They established that this conjugation of

CpG to the surface of tumor cells improves the uptake of tumor cell vaccines by Antigen Presenting Cells (APCs) and further triggers the functional maturation of the APCs.

### 3.3 Risks of using CpG-ODN *in vivo*

Though many benefits of CpG-DNA have been described but the application of these CpG-DNA motifs do involve some risks *in vivo*. Few toxic effects of CpG DNA were indicated such as it may trigger autoimmune diseases like Systemic lupus erythematosus<sup>85</sup>. Authors cautioned that excessive activation of immune system can cause systemic inflammatory response syndrome. A toxicity study of probiotic DNA was done on Swiss Albino Mice by Randhawa et al.<sup>38</sup> and it was concluded that toxicity is dose dependent. It is safe to administer 50-75µg/ml Probiotic DNA *in vivo*, but 100µg/ml and above dose lead to loss of hair and more aggressive behavior.

### 4. Future Recommendations

The efficacy of DNA can also be enhanced by loading it onto nanoparticles before introducing it in the body. So, nanoparticle assisted delivery may be a promising approach to alleviate the problem of instability and degradation of DNA and the studies over the nanoparticle approach for DNA delivery are going on.

### REFERENCES

1. McFarland LV, Meta-analysis of probiotics for the prevention of antibiotic associated diarrhea and the treatment of *Clostridium difficile* disease. *Am J Gastroenterol.* 2009; 101: 812–822.
2. Bengmark S and Gill GV, Probiotics potentiate IL-6 production in IL-1 beta-treated enterocytes through a heat shock-dependent mechanism. *Journal of the American College of Surgeons*, 2006; 203(3): S28-30.
3. Dahlqvist A and Gryboski JD, Inability of human small-intestinal lactase to hydrolyze lactulose. *Biochim. biophys. Acta (Amst.)* 1995; 110, 635-636.
4. Rendondo-Lopez V, Cook RL, Sobel JD, Emerging role of *lactobacilli* in the control and maintenance of the vaginal bacterial microflora. *Rev. infectious Diseases.* 1990; 12: 856–872.
5. Martin HL, Richardson BA, et al, Vaginal lactobacilli microbial flora and risk of human immunodeficiency virus Type 1 and sexually transmitted disease acquisition. *J. Infectious disease.* 1999; 180: 1863–1868.
6. Koop-Hoolihan L, Prophylactic and therapeutic uses of probiotics: A rev. *J. of the Am. Dietetic Assoc.* 2001.
7. Wang KY, Li SN, et al, Effects of ingesting *Lactobacillus* and *Bifidobacterium* containing yogurt in subjects with colonized *Helicobacter pylori*. *Am. J. Clin. Nutr.* 2004; 80: 737–741.
8. Baharav E, Mor F, Halpern M, Weinberger A, *Lactobacillus* GG Bacteria Ameliorate Arthritis in Lewis Rats. *J. Nutr.* 2004; 134: 1964–1969.
9. Jae-Seon So, Choong-Gu Lee, et al, *Lactobacillus casei* potentiates induction of oral tolerance in experimental arthritis. *Mol. Immunol.* 2008; 46: 172–180.
10. Jae-Seon So, Min-Kyung Song, et al, *Lactobacillus casei* enhances type II collagen/glucosamine-mediated suppression of inflammatory responses in experimental osteoarthritis. *Life Sci.* 2011; 88(7–8): 358–366.
11. Wang C, Shoji H, et al, Effects of oral administration of *Bifidobacterium breve* on fecal lactic acid and short-chain fatty acids in low birth weight infants. *Journal of Pediatric Gastroenterology and Nutrition.* 2007; 44: 252–257.
12. Takeda K and Okumura K, Effects of a fermented milk drink containing *Lactobacillus casei* strain Shirota on the human NK-cell activity. *J. Nutr.* 2007; 137(2): 791–793.
13. He T, Priebe MG, et al, Effects of yogurt and *bifidobacteria* supplementation on the colonic microbiota in lactose-intolerant subjects. *J. Appl. Microbiol.* 2007; 104: 595–604.
14. Ghadimi D, Holst RF, et al, Effects of Probiotic bacteria and their genomic DNA on T<sub>H</sub>1/T<sub>H</sub>2- cytokine production by peripheral blood mononuclear cells (PMBCs) of healthy and allergic subjects. *Immunobiology.* 2008; 213: 677-692.
15. Toh Z.Q, Anzela A, Tang MLK, Licciardi PV, Probiotic therapy as a novel approach for allergic disease. *Respiratory Pharmacol.* 2012; 3(171): 1-14.

16. van Baarlen P, Troost FJ, et al, Differential NF-kappaB pathways induction by *Lactobacillus plantarum* in the duodenum of healthy humans correlating with immune tolerance. Proc. Natl. Acad. Sci. USA. 2009; 106: 2371–2376.
17. Leyer G. J, Li S, Mubasher ME, Reifer C, Ouwehand AC, Probiotic effects on cold and influenza-like symptom incidence and duration in children. Pediatr. 2009; 124: 172 – 179.
18. Lonnermark E, Friman V, et al, Intake of *Lactobacillus plantarum* reduces certain gastrointestinal symptoms during treatment with antibiotics. J. Clin. Gastroenterol. 2010; 44: 106–112.
19. Dong P, Yang Y, Wang WP, The role of intestinal *Bifidobacteria* on immune system development in young rats. Early Hum. Devt. 2010; 86: 51–58.
20. Le Leu RK, Hu Y, Brown IL, Woodman RJ, Young GP, Synbiotic intervention of *Bifidobacterium lactis* and resistant starch protects against colorectal cancer development in rats. Carcinog. 2010; 31: 246–251.
21. Ataie-Jafari A, Larijani B, Alavi MH, Tahbaz F, Cholesterol-lowering effect of probiotic yogurt in comparison with ordinary yogurt in mildly to moderately hypercholesterolemic subjects. Ann. Nutr. Metab. 2009; 54: 22–27.
22. Bhatia A, Kaur M, Randhawa MK, Sharma A, Singla R, The anti-cholesterolemic effect of encapsulated synbiotics. J. Microbiol. Biotech. Res. 2012a; 2(5): 747-749.
23. Bhatia A, Kaur G, Kaur M, Singla R, Coencapsulation of Synbiotics for the evaluation of *in vivo* antidiabetic activity. Adv. Appl. Sci. Res. 2012b; 3(5), 3020-3024.
24. McCabe L.R, Irwin R, Schaefer L, Britton RA, Probiotic use decreases intestinal inflammation and increases bone density in healthy male but not female mice. J. Cell. Physiol. 2013; 228(8): 1793–1798.
25. Shida K and Nanno M. Probiotics and immunology: separating the wheat from the chaff. Trends in Immunology. 2008; 29(11): 565-573.
26. Ménard S, Laharie D, et al, *Bifidobacterium breve* and *Streptococcus thermophilus* secretion products enhance T helper 1 immune response and intestinal barrier in mice. Exp. Biol. Med. (Maywood). 2005; 230(10): 749-56.
27. Chuang L, Wu KG, et al, Heat killed cells of *Lactobacilli* skew the immune response towards Th1 polarization in mouse spleenocytes and dendritic cell-treated T cells. J. Agr. Food Chem. 2007; 55(26): 11080-86.
28. Ou CC, Lin SL, Pasi JJ, Lin MY, Heat killed lactic acid bacteria enhance immunomodulatory potential by skewing the immune response towards Th1 polarization. J. Food Sci. 2011; 76(5): 260-267.
29. Aattouri N and Lemonnier D. Production of interferon induced by *Streptococcus thermophilus*: role of CD4+ and CD8+ lymphocytes. J. Nutr Biochem. 1997; 8: 25-31.
30. Amrouche T, Boutin Y, Fliss I, Effects of Bifidobacterial cytoplasm peptide and protein fractions on mouse lymphocyte proliferation and cytokine production. Food Agr. Immunol. 2006a; 17: 29-42.
31. Amrouche T, Boutin Y, Fliss I, Effects of Bifidobacterial cytoplasm, cell wall and exopolysaccharide on mouse lymphocyte proliferation and cytokine production. Int. Dairy J. 2006b; 16: 70-80.
32. Makino S, Ikegami S, Kano et al, Immunomodulatory effects of polysaccharides produced by *Lactobacillus delbrueckii* spp. *Bulgaricus* OLL1073R-1. J. Dairy Sci. 2006; 89: 2873-2881.
33. Iliev ID, Tohno M, et al, Immunostimulatory oligodeoxynucleotide containing TTTCGTTT motif from *Lactobacillus rhamnosus* GG DNA potentially suppresses OVA-specific IgE production in mice. Scand. J. Immunol. 2008; 67(4): 370-376.
34. Krieg AM, Yi AK, et al, CpG motifs in bacterial DNA trigger direct B-cell activation. Nature. 1995; 374(6522): 546-549.
35. Weeratna R, Millan CLB, Krieg AM, Davis HL, Reduction of antigen expression from DNA vaccines by coadministered oligodeoxynucleotides. Antisense Nucleic Acid Drug Dev. 1998; 8(4): 351-356.
36. Lebeer S, Vanderleyen J, Keersmaecker SCJD, Genes and molecules of *Lactobacillus* supporting probiotic action. Microbiol. Mol. Biol. Rev. 2008; 72(4): 728-764.

37. Ng SC, Hart AL, Kamm MA, Stagg AJ, Knight SC, Mechanisms of action of probiotics: Recent advances. *Inflamm. Bowel Dis.* 2009; 15(2): 300-310.
38. Randhawa MK, Bhatia A, Ali SA, Balgir PP, In vivo comparison of probiotic bacteria's with their DNA: As augments of immune efficacy. *Adv. Appl. Sci.* 2012; 3(2): 826-832.
39. Verna EC and Lucak S, Use of probiotics in gastrointestinal disorders: what to recommend? *Ther Adv Gastroenterol.* 2010; 3(5): 307-319.
40. Lammers K.M, Brigidi P, et al, Immunomodulatory effects of probiotic bacterial DNA: IL-1 and IL-10 response in human peripheral blood mononuclear cells. *FEMS Immunology and Medical Microbiology.* 2003; 38: 165-172.
41. Rachmilewitz D, Katakura K, Karmeli F, Toll-like receptor 9 signaling mediates the anti-inflammatory effects of probiotics in murine experimental colitis. *Gastroenterol.* 2004; 126: 520–8.
42. Jijon H, Backer J, Diaz H, DNA from probiotic bacteria modulates murine and human epithelial and immune function. *Gastroenterol.* 2004; 126: 1358–1373.
43. Katakura K, Lee J, et al, Toll-like receptor 9-induced type I IFN protects mice from experimental colitis. *J. Clin. Invest.* 2005; 115: 695– 702.
44. Takahashi N, Kitazawa H, et al, An immunostimulatory DNA sequence from a probiotic strain of *Bifidobacterium longum* inhibits IgE production *in vitro*. *FEMS Immunol. Med. Microbiol.* 2006b; 46: 461–469.
45. Takahashi N, Kitazawa H, et al, An immunostimulatory oligodeoxynucleotide from *Bifidobacterium longum* suppresses Th2 immune responses in a murine model. *Clinical and Experimental Immunology.* 2006a; 145: 130-138.
46. Medina M, Izquierdo E, Ennahar S, Sanz Y, Differential immunomodulatory properties of *Bifidobacterium longum* strains: relevance to probiotic selection and clinical applications. *Clin. Exp. Immunol.* 2007; 150(3): 531-538.
47. Zhong Y, Huang J, Tang W, Chen B, Cai W, Effects of probiotics, probiotic DNA and the CpG oligodeoxynucleotides on ovalbumin-sensitized Brown-Norway rats via TLR9/NF- $\kappa$ B pathway. *FEMS Immunol. Med. Microbiol.* 2012; 66(1): 71-82.
48. Tokunaga T, Yamamoto H, et al, Antitumor activity of dedoxyribonucleic acid fraction from *Mycobacterium bovis* BCGI. Isolation, physicochemical characterization and antitumor activity. *J. Natl. Cancer Inst.* 1984; 72(4): 955-962.
49. Krieg AM, Motifs: the active ingredient in bacterial extracts. *Nat. Med.* 2003; 9: 831-835.
50. Krieg AM and Vollmer J, Toll-like receptors 7, 8, and 9: linking innate immunity to autoimmunity. *Immunol. Rev.* 2007; 220-251.
51. Fonseca DE and Kline JN, Use of CpG oligonucleotides in treatment of asthma and allergic disease. *Adv. Drug Deliv. Rev.* 2009; 61: 256-262.
52. Henнге UR, Benninghoff B, Ruzicka T, Goos M, Topical immunomodulators progress towards treating inflammation, infection, and cancer. *Lancet Infect. Diseases.* 2001; 1(3): 189-98.
53. Olishovsky SV, Kozak VV, Yanish YV, Shlyakhovenk VA, Immunostimulatory CpG DNA in cancer vaccinotherapy. *Experimental Oncology.* 2003; 25: 83-92.
54. Agrawal S and Kandimalla ER, Medicinal chemistry and therapeutic potential of CpG DNA. *Trends in Mol. Med.* 2002; 8(3): 114-121.
55. Klinman DM, Immunotherapeutic uses of CpG oligodeoxynucleotides. *Nature Reviews Immunology.* 2004; 4: 249-259.
56. Krieg AM, Homan LL, Yi AE, Harty JT, CpG DNA Induces Sustained IL-12 Expression *In Vivo* and Resistance to *Listeria monocytogenes* challenge. *J. Immunol.* 1998; 161: 2428-2434.
57. Gomis S, Babiuk L, et al, Protection of Chickens against *Escherichia coli* Infections by DNA Containing CpG Motifs. *Infection and Immunity.* 2003; 71(2): 857–863.
58. Jørgensen JB, Johansen LH, Steiro K, Johansen AJ, CpG DNA induces protective antiviral immune responses in Atlantic salmon (*Salmo salar* L.) *Virology.* 2003; 77(21): 11471-9.

59. Huang LY, Ishii KJ, Akira S, Aliberti J, Golding B, Th1-Like Cytokine Induction by Heat-Killed *Brucella abortus* is Dependent on Triggering of TLR9. *The Journal of Immunology*. 2005; 175(6): 3964-3970.
60. Talati AJ, Kim HJ, Kim YI, Yi YI, Yi AK, Role of bacterial DNA in macrophage activation by group B streptococci. *Microb. Infect.* 2008; 10: 1106-1113.
61. Zhu Y, Fan H, Miao J, Zou S, Protective effect of CpG DNA against mastitis induced by *Escherichia coli* in a rat model. *Veter. J.* 2008; 175(3): 369-378.
62. Easton A, Haque A, et al, Combining Vaccination and Postexposure CpG Therapy Provides Optimal Protection Against Lethal Sepsis in a Biodefense Model of Human Melioidosis. *The J. of Infect. Dis.* 2011; 204: 636-44.
63. Judy B.M, Taylor K, et al, Prophylactic application of CpG oligonucleotides augments the early host response and confers protection in acute melioidosis. *Plos one*. 2012; 7(3): 341-76.
64. McCluskie MJ and Krieg AM, Enhancement of infectious disease vaccines through TLR9-dependent recognition of CpG DNA. *Curr Top Microbiol Immunol*. 2006; 311: 155-78.
65. Yi AK, Chang M, Peckham DW, Krieg AM, Ashman RF, CpG oligodeoxyribonucleotides rescue mature spleen B cells from spontaneous apoptosis and promote cell cycle. *J. Immunol*. 1999; 160: 5898-906.
66. Liu N, Ohnishi N, Ni L, Akira S, Bacon KB, CpG directly induces T-bet expression and inhibits IgG1 and IgE switching in B cells. *Nature. Immunol.* 2003; 4: 687-693.
67. He B, Qiao X, Cerutti A, CpG DNA induces IgG class switch DNA recombination by activating human B cells through an innate pathway that requires TLR9 and cooperates with IL-10. *Immunol.* 2004; 173(7): 4479-91.
68. Kojima Y, Xin KQ, et al, Adjuvant effect of multi-CpG motifs on an HIV-1 DNA vaccine. 2002; 20(23-24): 2857-2865.
69. Cooper CL, Davis HL, et al, CpG adjuvant improves hepatitis B virus vaccine seroprotection in antiretroviral – treated HIV – infected adults. *AIDS*. 2005; 19: 1473-1479.
70. Sugai T, Mori M, et al, CpG containing oligodeoxynucleotide acts as an efficient adjuvant, counterbalancing the Th1/Th2 immune response in diphtheria-tetanus-pertussis vaccine. *Vaccine*. 2005; 23(46-47): 5450-56.
71. Maurer T, Pournaras C, et al, Immunostimulatory CpG-DNA and PSA-peptide vaccination elicits profound cytotoxic T-cell responses. *Urol. Oncolo.* 2011; 31(7) 1395–1401.
72. Zhao F, Liu S, et al, CpG adjuvant enhances the mucosal immunogenicity and efficacy of a *Treponema pallidum* DNA vaccine in rabbits. *Hum Vaccin Immunother.* 2013; 9(4): 753-60.
73. Wang YC, Tsai HP, et al, CpG DNA Is a Potent Enhancer of Humoral and Cell-Mediated Immune Responses against *Orientia tsutsugamushi* in C3H/HeN Mice. *J. Med. Sci.* 2013; 33(5): 233-243.
74. Horner AA, Widhopf GF, et al, ISS Immunostimulatory DNA inhibits IL-4-dependent IgE synthesis by human B cells. *J Allergy Clin Immunol*. 2001; 108: 417-23.
75. Kitagaki K, Businga T, Kline JN, Oral administration of CpG-ODNs suppresses antigen induced asthma in mice. *Clin. Experiment. Immunol.* 2005; 143(2): 249-259.
76. Suzuki M, Ohta N, et al, Immunotherapy with CpG DNA conjugated with T-cell epitope peptide of an allergenic Cry j 2 protein is useful for control of allergic conditions in mice. *Inter. Immunopharmacol.* 2007; 7(1): 46-54.
77. Xu W, Tamura T, Takatsu K, CpG ODN mediated prevention from ovalbumin induced anaphylaxis in mouse through B cell Pathway. *Inter. Immunopharmacol.* 2008; 8(2): 351-361.
78. Ashino S, Wakita D, et al, CpG-ODN inhibits airway inflammation at effector phase through down-regulation of antigen-specific Th2-cell migration into lung. *Int. Immunol.* 2008; 20: 259–266.
79. Miyazaki D, Kuo CH, Tominaga T, Inoue Y, Ono SJ, Regulatory Function of CpG-Activated B Cells in Late-Phase Experimental Allergic Conjunctivitis. *Investigative Ophthalmology & Visual Science*. 2009; 50(4): 1626-35.

80. Farrokhi S, Mousavi T, et al, Co-Administration of *Chenopodium album* Allergens and CpG Oligodeoxynucleotides Effects on Peripheral Blood Mononuclear Cells of Patients with Allergic Rhinitis Treated with Intranasal Corticosteroids and Antihistamines. Iran J Allergy Asthma Immunol. 2011; 10(2): 101-110.
81. Kaburaki Y, Fujimra T, et al, Induction of Th1 immune responses to Japanese cedar pollen (Cry j 1) in mice immunized with Cry j 1 conjugated with CpG oligodeoxynucleotide. Comparat. Immunol. Microbiol. Infect. Dis. 2011; 34(2): 157-161.
82. Wooldridge JE, and Weiner GJ, CpG DNA and cancer immunotherapy: orchestrating the antitumor immune response. Curr Opin Oncol. 2003; 15(6): 440-5.
83. Cornet S, Jamet JM, Lemonnier F, Kosmatopoulus K, Miconnet I, CpG oligonucleotides activate dendritic cells in vivo and induce a functional and protective vaccine immunity against a TERT derived modified cryptic MHC Class I- restricted epitope. Vacc. 2006; 24(11): 18880-1888.
84. Shirota H and Klinman DM CpG-conjugated apoptotic tumor cells elicit potent tumor-specific Immunity. Cancer Immunol Immunother. 2011; 60: 659–669.
85. Krieg AM, CpG DNA - A pathogenic factor in systemic lupus erythematosus. J Clin Immunol. 1995; 15: 284–92.